

#### **Introduction:**

The Curriculum development plays a very vital role in the development of quality of education. The education system should be such that students will be able to compete locally, regionally, nationally as well as globally. The present situation demands developing "learner-centric approach while redesigning of curriculum. There is also need to allow the flexibility in education system. The choice based credit system (CBCS) allows students to choose inter-disciplinary, intra-disciplinary courses, skill oriented papers and thus offers more flexibility for student. The courses can be evaluated following the grading system, which is considered to be better than the conventional marks system. In view of this,PAH Solapur University, Solapur has implemented Choice Based Credit System of Evaluation at Undergraduate level. While designing the syllabi of microbiology for undergraduate course for semester V and VI, an attempt has been made to follow the pattern given in the UGCs Undergraduate Template. This will help to bring a match across all the Indian universities.

Microbiology deals with the study of microorganisms. This branch of life science has immensely grown up widening its horizons and opening new frontiers of knowledge. The scope of microbiology as a subject is immense due to its ability to control all critical points of many fields like medical, dairy, pharmaceutical, industrial, clinical, research, water industry, agriculture, nanotechnology, etc. A career in microbiology is lucrative option. There is demand of trained microbiologists in a vast range of industries and institutes like research and development laboratories of government and private hospitals, research organizations, pharmaceutical, food, beverage and chemical industries. То the needs. discipline specific cater papers on industrial, agricultural, environmental, medical microbiology, microbial

biochemistry, virology, immunology have been included in the curriculum for semester V and VI. At the same time, the framework is so designed as to maintain standards of microbiology degree and the learning outcomes.

#### Learning Outcomes based approach to Curriculum Planning:

The Learning Outcomes-based Curriculum Framework (LOCF) for the B.Sc. (Honours) degree in Microbiology is designed to suit the need of the hour, in keeping with the emergence of new areas of microbiology. The framework is architected to allow for flexibility in programme design and course content development, while at the same time maintaining a basic uniformity in structure in comparison with other universities across the country. The programme is designed to build a strong microbiology knowledge base in the student and furthermore, acquaints the students with the applied aspects of this fascinating discipline as well. The student is thus equipped to pursue higher studies in an institution of her/his choice, and to apply the skills learnt in the programme to solving practical societal problems. The programme offers an elective course to the student for skill enhancement courses that prepares the student for an eventual job in academia or industry.

## Graduate Attributes in Microbiology:

Some of the characteristic attributes of an Honors graduate in Microbiology include:

Disciplinary Knowledge acquisition: gathers in-depth knowledge of basic and applied areas of microbiology.

- Core microbiology laboratory skills: understands various methods of safe handling, culturing and storage of microorganisms in the laboratory.
- Interdisciplinary approach: becomes aware of the role of microbiology in interdisciplinary research as well as in daily life.
- Environmental literacy: develops a basic understanding of the microbiological principles that have environmental implications, and gains an awareness of regulatory requirements and their compliance in biotechnology and microbiological research.
- > Thinking ability:inculcates independent thinking and apply knowledge acquired.
- Spirit of Team work: Reveals the importance of interacting with and working alongside people from diverse backgrounds.
- Global perspective: becomes acquainted with standard international practices and emerging technologies used to study microbes.
- Skills for Communication: acquires oral as well as written skills through oral presentations of ongoing developments in the field and compiling of information in brief in written format.
- > Ethical awareness: develops attitude of conducting their work with honesty
- > Self-motivation: develops planning, organization and time management skills.
- Digitally literatracy: : acquires Skills of using computers for Bioinformatics and computation and appropriate software for analysis of genomics and proteomics data, and employing modern bioinformatics search tools to locate, retrieve, and evaluate location and biological annotation genes of different species.

## **Outline of Choice Based Credit System:**

1. **Core Course**: A course, which should compulsorily be studied by a candidate as a core requirement is termed as a Core course.

2. Elective Course: Generally a course which can be chosen from a pool of courses and which may be very specific or specialized or advanced or supportive to the discipline/ subject of study or which provides an extended scope or which enables an exposure to some other discipline/subject/domain or nurtures the candidate's proficiency/skill is called an Elective Course.

Discipline Specific Elective (DSE) Course: Elective courses may be offered by the main discipline/subject of study is referred to as Discipline Specific Elective.

3. Ability Enhancement Courses (AEC): The Ability Enhancement (AE) Courses may be of two kinds: Ability Enhancement Compulsory Courses (AECC) and Skill Enhancement Courses (SEC). "AECC" courses are the courses based upon the content that leads to Knowledge enhancement; (i) Environmental Science and (ii) English/MIL Communication. These are mandatory for all disciplines. SEC courses are value-based and/or skill-based and are aimed at providing hands-on-training, competencies, skills, etc.

• **Credit**: Credit is a numerical value that indicates students work load (Lectures, Lab work, Seminar, Tutorials, Field work etc.) to complete a course unit. In most of the universities 15 contact hours constitute one credit. The contact hours are transformed into credits. Moreover, the grading system of evaluation is introduced for B.Sc. course wherein process of Continuous Internal Evaluation is ensured. The candidate has to appear for Internal Evaluation of 20 marks and University Evaluation for 80 marks.

# PAH Solapur University, Solapur Faculty of Science-New Choice Based Credit System (CBCS)- (w.e.f.2021-22) <u>Draft Structure for B. Sc-III</u>

Subject/ Name Core		and Type of the Paper	No. c papers/	fHrs/	wee	k	Total Marks	UA	CA	Credits
Course	Туре	Name	Practical	L	Т	Р	Per Paper			
Class :	<b>B.Sc</b> ]	III Semester – V								
Ability Enhancemer Course(AEC	nt CC)	English (Business English)	Paper- III	4.0			100	80	20	4.0
Discipline Specific Elective (DSE) (Students can op any one subjects among the three Subjects excluding interdisciplinary offered at B.Sc II.		<b>DSE-1A-</b> Virology	Paper- IX	3			100	80	20	4.0
		<b>DSE- 2 A-</b> Agricultural Microbiology	Paper -X	3			100	80	20	4.0
		DSE- 3 A- Immunology	Paper- XI	3			100	80	20	4.0
		<b>DSE 4 A-</b> : Industrial Microbiology	Paper- XII	3			100	80	20	4.0
		(Add-on-self learning)- MOOC/SWAYAM COURSE/INTERNSHIP								2.0
Grand Total				16.0			500	400	110	22
	B.Sc ]	III Semester –VI							•	
Ability Enhancement Course(AECC)		English (Business English)	Paper IV	4.0			100	80	20	4.0
any one	an opt	Genetics	Paper -XIII	3.0			100	80	20	4.0
subjects amo three Subjects ex interdisciplin offered at H	cluding ary	DEE 2D Mismahial	Paper- XIV	3.0			100	80	20	4.0

	<b>DSE- 3B-</b> Clinical Microbiology	Paper- XV	3.0	 	100	80	20	4.0
	<b>DSE 4B-</b> Environmental Microbiology	Paper- XVI	3.0	 	100	80	20	4.0
	SEC-							
Total (Theory)	<u> </u>		16.0	 	500	400	100	20
DSE - Practical	DSE- 1 A&B	Practical- IX & XIII		 5	100	80	20	4.0
(Annual Exam)	DSE -2 A&B	Practical- X&XIV		 5	100	80	20	4.0
	DSE- 3 A&B	Practical- XI&XV		5	100	80	20	4.0
	DSE- 4 A& B	Practical- XII & XVI		5	100	80	20	4.0
Total (Practicals)				20	400	320	80	16
Grand Total			32.0	20	1400	1120	280	58

# Summary of the Structure of B.Sc.Programme as per CBCS pattern

Class	Semester	Marks- Theory	Credits- Theory	Marks- Practical	Credits- Practicals	Total – credits
B.ScI	Ι	500	20			20
	II	550	20	400	16	36
B.ScII	III	350	14			14
	IV	350	14	300	12	26
B.ScIII	V	500	22			22
	VI	500	20	400	16	36
Total		2750	110	1100	44	154

<b>B.Sc.Programme :</b> Total Marks : Theory + Practical's = 2750 +1100 = 3950		
<b>Credits</b> : Theory + Practical's = $110 + 44 = 154$		
Numbers of Papers Theory: Ability Enhancement Course(AECC)		: 05
Theory: Discipline Specific Elective Paper (DSE)	: 08	
Theory: DSC : 12		
Skill Enhancement Courses /Add on	:01	
<b>Total</b> : Theory Papers	: 31	
: Practical Papers	:11	
Abbreviations :		
L: Lectures		
T: Tutorials		
P: Practicals		
UA : University Assessment		
CA : College Assessment		
DSC / CC: Core Course		
AEC : Ability Enhancement Course		
DSE : Discipline Specific Elective Paper		

SEC : Skill Enhancement Course GE : Generic Elective CA: Continuous Assessment ESE: End Semester Examination

## PAH SOLAPUR UNIVERSITY, SOLAPUR Faculty of Science Choice Based Credit System (CBCS) (W.e.f. 2021-22)

• Title of the Course:	B.Sc. Part-III (Honors)
• Subject:	Microbiology

• **Introduction**: This course provides a broad overview of Microbiology and to produces expert hands that would have sufficient knowledge and expertise to solve the urgent problems of the region by using Microbiology knowledge. The course structure is basic science centric where students learn core science and are taught necessary fundamental subject for that purpose.

• Objectives of the course: The objectives of B. Sc. Microbiology course are:

1) To impart knowledge with respect to the subject and its practicable applicability.

- 2) To enhance understanding of basic and advanced concepts in microbiology.
- 3) To develop the awareness of various emerging areas of Microbiology.
- 4) To train students for further studies helping in their bright career in the subject
- 5) To expose the students to different processes used in industries and in research field
- 6) To develop their ability to apply the knowledge of microbiology in day to day life.
- 7) To prepare the students to accept the challenges in life sciences.
- 8) To make students skillful to work in various industries, research labs and health sector.

**Course outcome and Advantages**: After completing the course students will be familiarized the with necessary laboratory techniques and tools of microbiology and provide an exposure in research, analytical and presentational skills. Microbiology has tremendous job potential. The successful students will be able well trained to get various microbiology related job.

- Medium of Instruction: English
- Syllabus Structure:
- The University follows semester system.
- An academic year shall consist of two semesters.
- B.Sc. Part-III Mcrobiology shall consist of two semesters: Semester V and Semester VI

<u>In semester V</u>: there will be Four DSC papers having paper IX to XII of 100 marks each. There will a Compulsory paper on "Ability Enhancement Compulsory Course (AECC)" on English and one self learning compulsory course of any one from -MOOC/SWAYAM COURSE/INTERNSHIP

<u>In Semester VI</u>: there will be four DSC papers having paper XIII to paperXVI of 100 marks each. There will a Compulsory paper on "Ability Enhancement Compulsory Course (AECC)" on English

The scheme of evaluation of performance of candidates shall be based on University Assessment (UA) as well as College Internal Assessment (CA) as given below.

For B.Sc.Part-III Microbiology SemV& VI the "internal assessment" will be based on Internal tests, Home assignment, Tutorials, Open Book Examination, Seminars, Group discussion, Brain storming sessions etc. as given below.

• Practical course examination is of 100 marks shall be conducted at the end of semester II. The practical examination of 400 marks shall also consist of 320 marks for University practical assessment and 80 marks for college internal assessment (CA).

• Scheme of Evaluation: As per the norms of the grading system of evaluation, out of 100 marks, the candidate has to appear for college internal assessment of 20 marks and external evaluation (University assessment) of 80 marks.

Semester – V: Theory: (100 marks): Comprising DSE-

a) University Examination (UA) (80 marks): No. of theory papers: 4 (paper IX to paper XII)

b) Internal Continuous Assessment (CA) (20 marks) No. of theory papers: 4 (paper IX to paper XII)

c) Compulsory paper on "Ability Enhancement Compulsory Course (AECC)" on English

d) One Add-on - self learning courseMOOC/SWAYAM COURSE/INTERNSHIP

Internal test- Home assignment / tutorials / seminars / viva/ group discussion/ outreach programs.

Semester - VI: Theory: (100 marks): Comprising DSE-

a) University Examination (UA) (80 marks): No. of theory papers: 4 (paper XIII to paper XVI)

b) Internal Continuous Assessment (CA) (20 marks) No. of theory papers: 4 (paper XIII to paper XVI)

Internal test- Home assignment / tutorials / seminars / viva/ group discussion/ outreach programs. **Practical Examination**: (400 marks)

:(80 UA + 20 CA))

:(80 UA + 20 CA)

University Examination (320 marks): Number of practicals : 04

- Practical-I: Based on Papers- IX & XIII (80 UA + 20 CA) Practical-II: Based on Papers- X & XIV :(80 UA + 20 CA)
- Practical-III: Based on papers -XI& XV

Practical-IV: Based on papers -XII& XVI

Internal Continuous Assessment: Total 80

(a) Internal practical test and

(b) Viva/group discussion/model or chart/attitude/attendance/overall behavior

(c) University practical examination of 320 marks (Practical I to IV for Four separate days) will be conducted at the end of semester VI

## Passing Standard:

The student has to secure a minimum of 4.0 grade points (Grade C) in each paper.

A student who secure less than 4.0 grade point (39% or less marks, Grade FC/FR) will be declared fail in that paper and shall be required to reappear for respective paper. A student who failed in University Examination (theory) and passed in internal assessment of a same paper shall be given FC Grade. Such student will have to reappear for University Examination only. A student who fails in internal assessment and passed in University examination (theory) shall be given FR Grade. Such student will have to reappear for both University examination as well as internal assessment. In case of Annual pattern/old semester pattern students/candidates from the mark scheme the candidates shall appear for the same stipulated marks of external examination and his/her performance shall be scaled to 100 marks.

## • ATKT:

passed in all papers except 6 (six) papers combined together of semester III and IV of B.Sc. Part-II Mcrobiology examination and clearly passed in B.Sc. Part-I-Mcrobiology shall be permitted to enter upon the course of Semester V of B.Sc. III Microbiology.

PAH SOLAPUR UNIVERSITY, SOLAPUR Faculty of Science and technology										
New CBCS Structure for B.Sc – III Microbiology Theory -										
Semester V						•				
Paper No.	Title of Paper	Hrs	/W	eek	Paper	UA	CA	Credits		
		L	Т	Р	Marks					
Ability	English	4	-	-	100	80	20	4		
Enhancement	(Business English)									
Course(AECC)										
DSE-IX	<b>DSE- 1A-</b> Virology	3	-	-	100	80	20	4		
DSE-X	<b>DSE- 2A-</b> Agricultural Microbiology	3	-	-	100	80	20	4		
DSE-XI	DSE- 3A- Immunology	3	-	-	100	80	20	4		
DSE-XII	<b>DSE 4A-</b> Industrial Microbiology	3	-	-	100	80	20	4		
SEC-	Add-on-self learning - MOOC/SWAYAM COURSE/INTERNSHIP		-					2.0		
Total		16	-	-	500	400	100	22		

Semester –VI
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Paper No.	Title of Paper		/Weel	k	Paper	UA	CA	Credit
-	-	L	Т	Р	Marks			S
Ability	English	4	-	-	100	80	20	4
Enhancement	(Business English)							
Course(AECC)								
DSEXIII	<b>DSE- 1B-</b> Microbial	3	-	-	100	80	20	4
	Genetics							
DSE-XIV	<b>DSE- 2B-</b> Environmental	3	-	-	100	80	20	4
	Microbiology							
DSE-XV	DSE- 3B- Clinical	3	-	-	100	80	20	4
	microbiology							
	Microbiology							
DSE-A	DSE 4B- Microbial	3	-	-	100	80	20	4
	Biochemistry							
SEC-								
Total		16			500	400	100	20

Practical No.	Paper No. based	Title of Paper		s/W	'eek	Paper Marks	UA	CA	Credit s
	on			Т	Р	-			
I	Papers - IX & XIII	DSE-1A- Virology AND DSE- 1B- Microbial Genetics	-	-	5	100	80	20	4
п	Papers - X&XI V	DSE-2A - Agricultural Microbiology AND DSE-2B - Environmental Microbiology	-	-	5	100	80	20	4
ш	Papers - XI&X V	DSE- 3A- Immunology AND DSE- 3 B - Clinical Microbiology	-	-	5	100	80	20	4
IV	Papers - XII & XVI	DSE 4 A- : Industrial Microbiology AND DSE 4B- Microbial Biochemistry	-	-	5	100	80	20	4
	Total		-	-	20	400	320	80	16

#### **Abbreviations:**

- L: Lectures
- T: Tutorials

P: Practicals

- UA:
- CA:
- UA:
- University Assessment by End Semester Examination College Assessment by Internal Continuous Examination University Assessment: University Theory paper shall be of 70 marks College Assessment: The internal examination for theory and practical course CA:

# [Credits -4, Total Lectures-60]

# DSE – 1- A: Paper MIC IX: Virology

Unit I Introduction and Classification of Viruses	(11)
A. General properties and structure of virus	
B. Viroids and Prions	
C. Viral classification on the basis of LHT system and as per international committ	ee
Unit II Reproduction of bacterial viruses	(11)
A. T <sub>4</sub> Bacteriophage - Lytic cycle	
B. Temperate phages and lysogeny of $\lambda$ phages	
Unit III Animal Viruses	(14)
A. Reproduction of Animal viruses: Adeno viruses and Influenza viruses	
B. Oncogenic Viruses- i) Types of Oncogenic viruses, DNA and RNA viruses, (ii)	
Types of cancer, Characteristics of cancerous cells, (iii) Hypotheses of Cancer:	
Somatic mutation, Viral gene and Defective immunity	
Unit IV Plant Viruses	(12)
A. Viral plant Diseases – Tobacco mosaic virus[TMV], Cauliflower Mosiac virus[	CaMV]
B. Prevention and Control of Plant Viral Diseases	
Unit V Techniques in Virology	(12)
A. Isolation, cultivation, Purification and Enumeration of viruses	(12)
<b>B.</b> One step growth experiment	
References:	
1. General microbiology – Stanier	
2. General microbiology – Pawar and DaginawalaVol I and II	
3. Genetics of bacteria and their viruses – William Hays	
4. Virology – Biswas	
5. Virology – Luria	
6. Microbiology - Prescott, Harley and Klein's, Willey Sherwood	
Woolverton,McGraw – Hill International Edition, (2008).	
7. Plant viruses- by Mathews	

- 8. Microbiology by Davis
- 9. Plant diseases by Singh

# [Credits -4, Total Lectures-60]

# DSE – 2 – A: Paper MIC - X: Agricultural Microbiology

Unit I- Introduction toSoilMicrobiology	(11)
<ul> <li>A. Introduction to soil– Definition of soil, Soil formation, types, structure and properties</li> <li>B. Soil as anecosystem, rhizosphere and phyllosphere</li> <li>C. Soil microorganisms, types and theirrolein soil fertility, humus.</li> </ul>	
D. Interactions in soil.	
Unit II- Role of microorganisms inelementalcycle	(11)
A. Carboncycle B. Nitrogencycle	
C. Sulphurcycle	
D. Phosphorouscycle	
Unit III- Composting and Biodegradation	(14)
A. Compost production with reference to organic waste, types of microorganisms, and factors affecting - aeration, C:N:P ratio, moisture content, temperature, pH, and period	
of composting.	
1. Greenmanure	
<ol> <li>Farm yardmanure</li> <li>Towncompost</li> </ol>	
4. Vermicompost	
B. Biodegradation of Cellulose, hemicelluloses, Lignin, Pectin, and Pesticides	
Unit IV-Plantpathology	(12)
A. Common symptoms produced by plantpathogens	
B. Modes of transmission of Plant diseases: Oily spots on pomegranate –	
<i>Xanthomonasoxynopodis</i> , whip smut of sugarcane, soft rot ofpotato C. Control measures of plantdiseases	
Unit V Applications of BiotechnologyinAgriculture	(12)
<ul> <li>A. Biofertilisers (Azo and Rhizo and PSB) production and applications.</li> <li>B. Bioinsecticides – Bacillus thuriengenesis and Trichodermaviridae, Other examples</li> </ul>	
C. Genetically Modified Crops with examples	
D. Viral pesticides:Concepts & applications	

## References

- 1. Soil Microbiology Subbarao, N.S.
- 2. Microbial dynamics and diversity DesyStaley

- 3. Biology of Microorganisms Brock, Parker, Madigen, 9thedition
- 4. Agricultural Microbiology- Bagyaraj and Ghosh
- 5. Plant Diseases- SinghR.S.
- 6. Soil Microbiology Alexander.
- 7. Industrial Microbiology PatelA.H.
- 8. Textbook of Biotechnology R.C.Dubey,

[Credits -4, Total Lectures-60]

#### DSE - 3 - A: Paper MIC - XI: Immunology

#### Unit I. Immune system and Adaptive Immunity

(16)

## A] Immune system

a]structure &function of lymphoid organs:

1]Primary and organs - Thymus, bone marrow,

2] Secondary lymphoid organs-- spleen , lymph node and Mucosa associated lymphoid tissue(MALT)

b]cells of immune system &their role

i]. Classification of cells of immune system-Lymphoid and myeloid cells

ii]. Structure and functions of Lymphoid cells- T cells and T cell subsets, NK cells, B cells and dendritic cells

iii]. Structure and functions of myeloid cells – Granulocytes, Monocytes and macrophages

## **B**]Adaptive Immunity

-	-			
a) Humoral	(antibody) mediated	d response –Cells	involved and	l mechanism
	(			

b) Cell mediated – cells involved, mechanism (endogenous and exogenous pathways); cytokines and their role

c) Primary and secondary immune response

d) Monoclonal antibodies – i) Production (hybridoma technology) ii)

Applications of Monoclonal antibodies in Diagnosis, Research and Treatment

## Unit II.Major Histocompatibility complex

A. Organization of MHC genes in man, Classes of MHC molecules – structure and their role,

B. HLA typing

C. Types of grafts and their rejection.

## Unit III.Complement system

A. Components of complement and their properties,

B. Activation of complement -classical and alternate pathway

C. Biological effects of complement

## Unit IV Immunological disorders

- A. Hypersensitivity Classification based on
  - 1. Time:Immediate and delayed typehypersensitivity
  - 2. Mechanism of Pathogenesis
    - a) Type I : Anaphylaxis, Atopy
    - b) Type II : Autohaemolytic anemia
    - c) **Type III :** Arthus reaction, Serum sickness
    - d) Type IV: contact dermatitis, Allergy of infection

# B. Autoimmunity :a. Mechanism of Autoimmunity

- b. Types of Autoimmune diseases-
- a) Hemocytolytic b) Organ Specific(Graves disease, Myasthenia gravis, pernicious anemia)

(12)

(06)

(16)

#### Unit V Immunohaematology

- A. ABO blood group system
- B. Rh blood group system
- C. Blood transfusion reaction and its complications

#### References

- 1. Essentials of Immunology Roitt Evan, Brostoff J. Male D. (1993) 6th Edition.
- 2. Immunology Kuby J. (1996) W.H. Freeman and Co.
- 3. Immunology Fudenberg
- 4. Medical Microbiology Davis and Dulbecco
- 5. Medical laboratory technology RamnaikSood
- 6. Diagnostic Microbiology Bailey's and Scotts
- 7. Immunology a problem approach by Wood, Hood and Weison
- 8. Medical Bacteriology Dey and Dey
- 9. Handbook of Immunology- G.P. Talwar (1983) Vikas Publishing Pvt.
- Ltd 10.Textbooks of medical microbiology-Anant Narayan
- 11. Immunolgy& Serology-Carpenter.

#### [Credits -4, Total Lectures-60]

DSC – 4A : Paper MIC – XII : Industrial Microbiology	
Unit I Food Microbiology	(11)
A. Food as a substrate for microorganisms	
B. Food Spoilage (Meat and Poultry, Fruits and Vegetables)& food borne diseases-food	
infection (Salmonella) & food poisoning (Clostridium)	
C. Principle and methods of food preservation D. Food Fermentations – i) Idli ii) Bread	
D. Food Permentations – 1) Idn II) Bread	
Unit II Dairy Microbiology	(11)
A]Spoilage of milk	
B]Fermented dairy products: i) Cheese ii) Yogurt	
Unit III Industrial production of	(12)
A. Streptomycin	
B. Lysine	
C. rDNA products – Insulin	
D. Vitamin B12	
Unit IV Production of alcoholic beverages	(12)
C	(12)
A. Grape wine – Definition, types, production of White table wine and Red table wine B. Post fermentation spoilage of wines: Microbial and non microbial spoilage of wines	
C. Beer – Definition, types, production of Lager beer and Ales Beer.	
C. Deel Dermition, Opes, production of Luger even and thes Deen	
Unit V Downstream processing and quality control:	(14)
A) Downstream processing: Filtration, Cross flow filtration, Flocculation, Whole broth	
processing Solvent extraction, Concentration, Centrifugation,	
Crystallization, Distillation, Adsorption elution, Precipitation and	
Chromatography	
<b>B</b> ) Quality control in fermentation industry: Test for sterility, pyrogenicity,	
allergy, Carcinogenicity, toxicity for Pharmaceutical and health care and food	
products	
References	
1. Principles of fermentation technology – Whitkar and Stanbury	

- 2. Pharmaceutical Microbiology Huggo
- Biochemistry Fox and Nelson
   Industrial Microbiology Prescott and Dunn
   Microbial technology Peppler
- 6. Advances in Biotechnology S.W. Jogdand.
- 7. Textbook of Biotechnology R.C. Dubey,
- 8. Biotechnology B.D. Singh

9. Industrial Microbiology – Casida
10. Industrial Microbiology by A.H. Patel.
11. Food Microbiology: an Introduction by Adam and Dick
12. Food Microbiology by Frazier

# [Credits -4, Total Lectures-60]

# DSE – 1B – B:PaperMIC - XIII: Microbial Genetics

<ul> <li>Unit I: Basic concepts of microbial genetics</li> <li>A) Structural organization of <i>Escherichia coli</i> chromosome, folded fiber model</li> <li>B) Replication of DNA: Enzymes involved and mechanisms of replication</li> <li>C) Transcription: RNA polymerase enzyme, process and post transcriptional modification</li> <li>D) Operon concept – Lac Operon</li> </ul>	(11)
<ul> <li>Unit II: Effect of mutation in bacteria</li> <li>A) Effect of mutation on translation</li> <li>B) Effect of mutation on phenotypes</li> <li>C) Time course of phenotypic expression</li> <li>D) Selection, detection and Isolation of mutants</li> <li>E) Genetic Complementation – Cis – Trans Test</li> </ul>	(12)
<ul> <li>Unit III: Genetic engineering and Protein engineering</li> <li>A) Introduction, Tools and Techniques of Genetic engineering</li> <li>B) Applications of Genetic engineering</li> <li>C) Protein Engineering – concept and applications</li> </ul>	(14)
<ul> <li>Unit IV : Techniques in molecular biology</li> <li>A) Electrophoresis of DNA.</li> <li>B) DNA sequencing – Sanger Dideoxy method</li> <li>C) DNA finger printing- method and applications</li> </ul>	(12)
<ul> <li>Unit V Bioinformatics</li> <li>A. Introduction to Bioinformatics.</li> <li>B. Introduction to major bioinformatics resources on Internet: National Centre for Biotechnology Information (NCBI), DDBJ, EMBL.</li> <li>C) Protein data bank (PDB) and Nucleic acid sequence database (GenBank)</li> <li>D) The Basic Local Alignment Search Tool (BLAST)</li> </ul>	(11)
References:	
1. General microbiology – Stanier	

- 2. General microbiology Pawar and DaginawalaVol I and II
- 3. Biochemistry Lehninger
- 4. Molecular Biology of Gene J.D. Watson
- 5. Recombinant DNA J.D. Watson

- 6. Microbiology Davis
- 7. Biochemistry Purohit
- 8. Genetics of bacteria and their viruses William Hays
- 9. http://www.ncbi.nlm.nih.gov/

[Credits -4, Total Lectures-60]

## DSE – 2 – B Paper MIC - XIV: Environmental Microbiology

#### Unit I: Air microbiology and biosafety issues

(16)

(10)

(8)

(14)

A]Concept and definition of biosaftey, objectives of biosafety, NIH guidelines

B] Regulatory framework of biosaftey in India-

a]Recombinant DNA, Advisory committee[RDAC]

b] Institutional biosaftey committee[ISBC]

c]State biosaftey coordination committee [SBCC]

d] District level biosaftey committee[DLBC]

C]Microorganisms in air – Launching, transport and deposition of aerosols, survival of microorganisms in air D]Significance of microorganisms in air (extramural and intramural) EMethods to study air borne microorganisms. Sampling, qualitative and quantitative methods. F]Bioaerosal control (ventilation, filtration, biocidal control, UV gaseous (quarantine)

#### Unit II : Marine microbiology and Fresh water ecosystem

A. Microorganisms in marine water, methods to study aquatic microorganisms. Characteristics of marine environment, types of organisms and their role.

B. Fresh water ecosystem : Eutrophication, Types of fresh water bodies a) Classification of lakes

b) Sources c) Consequences d) Control

#### **Unit III Extremophiles:**

General characteristics of extremophiles and their role -Acidophiles, Alkalophiles, Thermophiles, Psychrophiles, Barophiles and Osmophiles

#### Unit IV Environmental impact assessment and Industrial Waste Management: (12)

A) Types of wastes, Waste water assessment and management, BOD, COD,
B. Industrial waste treatment : Characteristics and treatment of wastes from different industries, – paper and pulp, sugar and distillery, textile, and dairy industries,
C) Bioremediation :Lead, mercury, arsenic and radioactive substances

## Unit V Geomicrobiology and carbon sequestration

A) Introduction, Microorganisms involved, Biochemistry of microbial leaching,

Commercial leaching – slope, heap, in situ leaching, Leaching of Iron, Copper and Uranium,

B) Oilrecovery: Methods – primary, secondary, and microbially enhanced oil recovery,

C) Concept of carbon sequestration and carbon credit

# **References:**

- 1. Physiology and Biochemistry of Extremophiles- Charles Gerday and Nicolas Glansdorff
- 2. Environmental Microbiology Maier
- 3. Microbial ecology Fundamentals and applications Atlas and Bartha
- 4. Microbial dynamics and diversity Desy Staley
- 5. Biology of Microorganisms Brock, Parker, Madigen, 9th edition
- 6. Microbiology Prescott and Harley, 5th edition

[Credits -4, Total Lectures-60]

## DSE - 3 B- Paper MIC - XV: Clinical Microbiology

#### **Unit I: Microbial diseases**

(characters of etiological agent, , modes of transmission, pathogenesis, symptoms, laboratory diagnosis, prophylaxis and treatment of following diseases)

**1.Bacterial**: 1. Pseudomonas aeruginosa 2.Mycobacterium leprae 3.Clostridium perfringens 4.Vibrio cholerae

**2.Viral:** 1. Rabies 2.AIDS 3.Swine flu 4. Ebola

3.Fungal:1.Dermatophycoses[Tinea],2.cryptococcosis

4.protozoal: 1.Malaria 2. Giardiasis

## **Unit II :Biomedical Waste Management**

Laboratory disposal of – Sharp devices, Smeared Slides, Cultures, Culture Media and Clinical Samples, Fomites

Unit III mechanism of Pathogenecity –

- **1.** Definition & Concept
- 2. Basic principles of Microbial adhesion
- 3. Mechanism of Bacterial invasion
- 4. Bacterial toxins Types & mechanism of action

5. Mechanism of pathogenecity of viral & fungal infections

#### Unit IV:Vaccinology & bioweapons

- 1. Basic concept
- 2. Types:A]traditional-live attenuated,killed,toxoids

B]New generation vaccines: subunit, Recombinant ,conjugated, DNA 3.Definition,Characteristics of bioweapons ,Advantages &disadvantages of bioweapons,Examples

#### **Unit-V : Chemotherapy**

- 1. Ideal characteristics of chemotherapeutic agents
- 2. Mechanism of action of different chemotherapeutic agents:

A]antibiotics

i. Acting on Cell Wall: Penicillin, Bacitracin, Vancomycin,

ii. Acting on Protein Synthesis: Streptomycin, Chloramphenicol,

iii. Acting on nucleic acid synthesis: quinolenes, rifampicin

iv. Drugs Acting on folic acid synthesis: Sulphonamide, Trimethoprim,

B]antiviral agents

C]antifungal agents

D]antiprotozoal agents

3.Mechanism of antibiotic resistance

4.Tests to guide chemotherapy:diffusion and broth dilution methods

(14)

(**06**) a and

(24)

(8)

(8)

## **References:**

- 1. Pharmaceutical Microbiology Huggo
- 2. Text book of Medical Microbiology Ananthnarayan
- 3. Review of Medical Microbiology Jawetz et al
- 4. Microbiology Zinsser
- 5. Medical Microbiology Cruickshank
- 6. Medical Microbiology Davis and Dulbecco
- 7. Parasitology Chattergii
- 8. Medical laboratory technology RamnaikSood
- 9. Diagnostic Microbiology Bailey's and Scotts
- 10. Medical Bacteriology Dey and Dey

[Credits -4, Total Lectures-60]

#### DSE – 4– B: Paper MIC - XVI: Microbial Biochemistry

# Unit I Enzyme, Enzyme kinetics and regulation (16)A) Enzymes: (i) Definition, properties, structure, specificity, mechanism of action (Lock and key model, induced fit hypothesis) (ii) Allosteric enzymes – Definition, Two models explaining mechanism of action (Sequential and Concerted) (iii) Ribozymes and Isozymes iv) Factors affecting catalytic efficiency of enzymes i) Proximity, orientation ii) Strain and distortion iii) Acid base catalysis iv) Covalentcatalysis B) Enzyme kinetics – Derivation of MichaelisMenten equation, Significance of Km and Vmax **C)** Regulation of enzyme synthesis i) Positive control – Arabinose Operon ii) End product repression – Tryptophan Operon iii) Catabolite repression Unit II Extraction, purification and assay of enzymes (10)A) Cell disruption and homogenization of membrane bound enzymes, Extraction B) Purification of enzymes on the basis of - i) Molecular size ii) Solubility iii) Electric charge iv) Adsorption characteristics) Biological affinity C) Immobilization of enzymes – Methods and applications D) Assay of enzymes (enzyme unit, enzyme activity, Specific activity) **Unit III Assimilation of:** (10)A. Carbon B. Nitrogen – N2 and NH3 (GOGAT) C. Sulphur **Unit IV Bioenergetics:** A) Pyruvate as key metabolite inCarbohydratemetabolism (14)B) Metabolic Pathways i) ED pathway ii) Glyoxylate bypass iii)Pentose Phosphate Pathway iv)Phosphoketolase pathway C) Bioluminescence

## **Unit VBiosynthesisof:**

A) Nucleotides- Purines and pyrimidines B) Protein C) Peptidoglycan

## **References:**

- 1. Molecular Biology of Gene J.D. Watson
- 2. Principles and techniques of Practical Biochemistry K. Wilsons J.Walkar.
- 3. Analytical Chemistry Robert B. Dilts
- 4. Chromatographic methods by Braithwaite and White
- 5. Outline of Biochemistry Cohn and Stump
- 6. Biochemistry West and Todd Russel
- 7. Biochemistry Lehninger
- 8. Enzymes Dixon and Web
- 9. Biological chemistry Mahler and Cordes
- 10. Nature of Enzymology R.L. Foster
- 11. Microbial technology Peppler
- 12. Biochemistry A problem approach by Wood, Hood and Weison

#### PAH SOLAPUR UNIVERSITY, SOLAPUR (CBCS) B.Sc. III-Microbiology w. e. f. June 2021 --Practicals Syllabus ( Credits-4 )

## PRACTICAL COURSE

#### **Practical V:**

1. Isolation of DNA from bacteria by J. Marmur's method

2. Electrophoretic separation of DNA

3. Isolation of coliphages from sewage

4. One step growth curve

5. Determination of dose of U.V. by UV survival curve

6. Isolation of Lac negative mutants of *E.coli* by visual detection method.

7. Isolation of Streptomycin resistant mutants by gradient plate technique.

8. Isolation of Vitamin B12 requiring (auxotrophic) mutants by replica plate technique.

9. Cultivation of Viruses in Embryonated chicken egg

10. Study of Virally infected lesions of Plant materials

11. Browsing of National Centre for Biotechnology Information (NCBI), DDBJ and EMBL websites.

12. Exploring protein sequence database (PDB) and GeneBank and BLAST.

#### **Practical VI:**

1. Isolation of Azotobacter from soil. (Identification up to genus level)

2. Isolation of *Rhizobium* from root nodules.

3. Isolation of *Xanthomonas* from infected plant material

4. SPC of market Biofertilizers.

5. Estimation of available phosphorous from soil (Stannous chloride method)

6. Estimation of Calcium and Magnesium from soil (EDTA method)

7. Determination of organic carbon contents of soil (Walkley and Black method)

8Microbiological analysis Drinking water: Presumptive, Confirmed and Completed test 9Determination of potability of water by MPN.

10Waste water analysis: Chemical Oxygen Demand

(COD)

11.Biological Oxygen Demand (BOD)

#### **Practical VII:**

- 1. Separation and Preservation of Serum and Plasma
- 2. Widal test (quantitative test), RA test, Pregnancy test
- 3. Haematology -
  - 1] Estimation of Hb by Sahlis method
  - 2] Total blood cell count:RBC count, WBC count,
  - 3]differential WBC count

4]Determination of Erythrocyte sedimentation Rate

4. Isolation of pathogen from clinical sample Pseudomonas aeruginosa/E.coli

5.Study of determination of effectiveness of antiseptic agents (- tincture iodine, 3% H2O2,

70% alcohol, 5% chlorine bleach) against selected organisms (Escherichia coli,

Staphylococcus aureus, Bacillus) by disc diffusion method
6.Antimicrobial susceptibility testing by disc diffusion
method
7.Urine analysis :
A]Microscopic examination – pus cells, RBc, bacteria, crystals.
B] Chemical examination –
1]glucose (benedict's method,
2]protein (acetic acid),
3]bile salt (sulphur method),
4]ketone bodies (Rothera's test)

## **Practical VIII:**

1. Examination of milk

i) DMC

**ii**) Quantitative analysis of milk by SPC (using nutrient agar)

2. Phosphatase Test (qualitative)

3. Isolaton and identification of microorganisms from spoiled food

4. Bioassay of Streptomycin

5. Estimation of alcohol by using K2Cr2O7

6.Diauxic growth curve of Escherichia coli (glucose and lactose).

7.Immobilization of enzyme by using Sodium alginate

8. Thin layer chromatography- amino acid

9. Study of Substrate concentration on enzyme activity.

10.Purification of enzyme and study of its activity

11.Production of citic acid by Aspergillus niger and estimation of Citric acid by titration method

12.Bioassay of Vitamin B12

13.Sterility testing of media and pharmaceutical

products

14..Isolation of Lactic acid Bacteria (MRS Medium)

15.Amylase assay (Iodometric method)

#### **Practical Examination**

A) The university practical examination will be conducted on four (4) consecutive days for not less than 6 hours on each day of the practical examination. The practical examination shall be conducted by the two external examiners appointed by the University.

B) Each candidate must produce a certificate from the Head of the Department in his/her college stating that he/she has completed in a satisfactory manner the practical course on the guidelines laid down from time to time by Academic council on the recommendation of Board of studies and has recorded his/her observations in the laboratory journal and written a report on each exercise performed. Every journal is to be checked and signed periodically by a member teaching staff and certified by the Head of the Department at the end of the year. Candidates are to produce their journal at the time of practical examination. Candidate has to visit two places of Microbiological interest (Pharmaceutical industry, Dairy, Research institutes etc) and submit the report of their visit at the time of examination. The report should be duly certified by the Head of the Department.

#### Distribution of Marks for practical examination

1) One major experiment: 30 marks

2) Two minor experiment: 15 marks each

3) Journal: 5 marks

4) Viva:10

#### **Total marks:**

Practical V:75Practical VI:75Practical VII:75Practical VIII:75Tour Report :20Total Marks:320

Practical V Marks: 75

#### **Que1. Major Experiments**

Isolation of DNA from bacteria by J. Marmur's method

OR

Isolation of coliphages from sewage

#### **Que2. Minor Experiments**

One step growth curve OR

Determination of dose of U.V. by UV survival curve

OR Cultivation of Viruses in Embryonated chicken egg

OR

Study of Virally infected lesions of Plant materials OR

Browsing of National Centre for Biotechnology Information (NCBI), DDBJ and EMBL websites

#### **Que3. Minor Experiments**

Isolation of Lac negative mutants of *E.coli* by visual detection method. OR Isolation of Streptomycin resistant mutants by gradient plate technique. OR Isolation of Vitamin B12 requiring (auxotrophic) mutants by replica plate technique. OR Exploring protein sequence database (PDB) and GeneBank and BLAST.

#### **Practical VI**

#### **Que 1 Major Experiment**

Isolation of Azotobacter from soil. (Identification up to genus level)

OR

Isolation of *Rhizobium* from root nodules.

OR

Isolation of *Xanthomonas* from infected plant material

OR Biological Oxygen Demand (BOD) OR

Microbiological analysis Drinking water: Presumptive, Confirmed and Completed test

#### Que2 Minor Experiment

SPC of market **Biofertilizers.** 

OR

Determination of potability of water by MPN.

#### **Que 3 Minor Experiment**

Estimation of available phosphorous from soil (Stannous chloride method)

OR

Estimation of Calcium and Magnesium from soil (EDTA method)

OR

Determination of organic carbon contents of soil (Walkley and Black method)

OR

Waste water analysis: Chemical Oxygen Demand (COD)

#### **Practical VII**

#### Que. 1 Major Experiment

Isolation of pathogen from clinical sample Pseudomonas aeruginosa/E.coli

**Que 2 Minor Experiment** Widal test (quantitative test)/ RA test,/Pregnancy test

OR Estimation of Hb by Sahlis method

OR RBC count/ WBC count/differential WBC count OR

Determination of Erythrocyte sedimentation Rate OR Microscopic examination – pus cells, RBC, bacteria, crystals. OR Chemical examination – glucose (benedict's method,protein (acetic acid),bile salt (sulphur method), ketone bodies (Rothera's test)

#### **Que. 3 Minor Experiment**

Study of determination of effectiveness of antiseptic agents (– tincture iodine, 3% H2O2, 70% alcohol, 5% chlorine bleach) against selected organisms (*Escherichia coli, Staphylococcus aureus, Bacillus*) by disc diffusion method

OR Antimicrobial susceptibility testing by disc diffusionmethod

#### **Practical VIII**

## Que 1 Major experiment

Bioassay of Streptomycin OR Bioassay of Vitamin B12

#### OR

Isolaton and identification of microorganisms from spoiled food

OR

Examination of milk -DMC & Quantitative analysis of milk by SPC (using nutrient agar)

#### OR

Isolation of Lactic acid Bacteria (MRS Medium

#### **Que 2 Minor Experiment**

Phosphatase Test(qualitative)

OR

Estimation of alcohol by using K2Cr2O7 OR Diauxic growth curve of Escherichia coli (glucose and lactose).

OR Sterility testing of media and pharmaceutical products OR Amylase assay (Iodometric method)

#### **Que 3Minor Experiment**

.Immobilization of enzyme by using Sodium alginate

## OR

Thin layer chromatography- amino acid

OR Study of Substrate concentration on enzyme activity.

# OR

Purification of enzyme and study of its activity OR Estimation of Citric acid by titration method

#### List of the Minimum equipments and related requirements for B.Sc - III

- 1) Replica plating units for genetics experiments: Two
- 2) Rotary shaker for fermentation experiments: One

3) Centrifuge (High speed): One

4) Hot plate: One

5) Hot air oven: One

6) Bacteriological incubator: One

7) Spectrophotometer: One

8) Research Microscope: one for each student

9) Haemocytometer: Two

10) Haemoglobinometer: Two

11) ESR stands and tubes: Two

12) Separate room for fine instruments of size 10'×15' feet dimension

13) A separate culture room of at least  $10' \times 10'$  feet dimension

14) Electrophoresis assembly: One

15) Laminar air flow cabinet: One

16) Distillation assembly: One (Glass)

17) Reflux assembly: Four

18) Serological water bath: One

19) Colony counter: One

20) Refrigerator: One

21) TLC UNIT: One

22) Hand Refractometer

23) Computer with Internet facilities and printer: One

24) Micropipette: One

25) Anaerobic Jar: One

26) Heating Mantle: One

27) UV Chamber